



Pergamon

Tetrahedron Letters 41 (2000) 3907–3910

TETRAHEDRON
LETTERS

Highly regio- and stereospecific hydroxylation of C-1 position of 2-deacetoxytaxinine J derivative with DMDO

Tohru Horiguchi, Qian Cheng and Takayuki Oritani *

Laboratory of Applied Bioorganic Chemistry, Graduate School of Agricultural Science, Tohoku University,
1-1 Tsutsumidori-Amamiyamachi, Aoba-ku, Sendai 981, Japan

Received 14 February 2000; revised 13 March 2000; accepted 24 March 2000

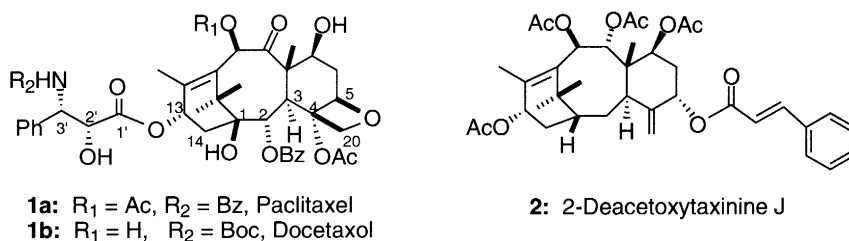
Abstract

A simple chemical oxidation of 2-deacetoxytaxinine J derivative **3** using an excess of dimethyldioxirane (DMDO) results in a highly regio- and stereospecific hydroxylation of the C-1 position to afford the 1 β -hydroxy α -4(20)-epoxide **6** and 1 β -hydroxy β -4(20)-epoxide **7**. A plausible mechanism of this reaction via ‘insertion’ is proposed. © 2000 Elsevier Science Ltd. All rights reserved.

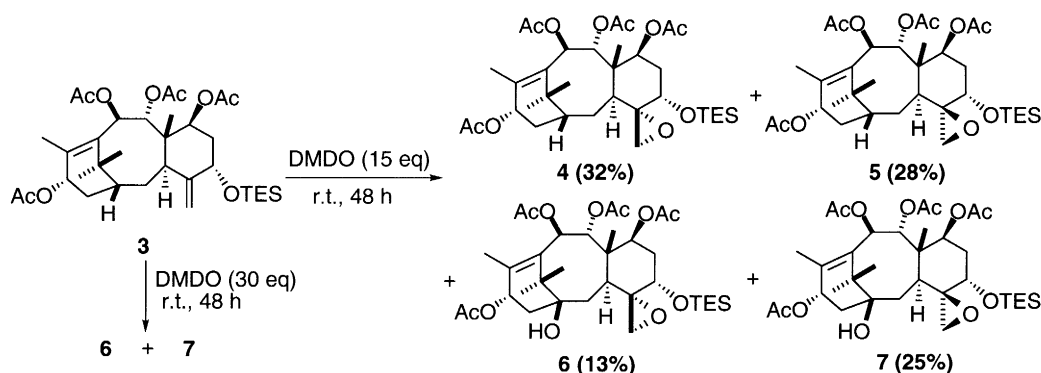
Keywords: taxoids; dioxiranes; hydroxylation; regioselectivity; stereospecificity; epoxides; epoxidation.

Currently, diterpenoid paclitaxel¹ (Taxol[®], **1a**) and docetaxel² (Taxotere[®], **1b**) are regarded as powerful therapeutic drugs for cancer chemotherapy. Both compounds exhibit potent antitumor activity against various cancers that have been uneffectively treated by existing chemotherapeutic drugs,^{2,3} and both have been approved by the FDA for the treatment of advanced ovarian cancer and breast cancer. Paclitaxel was also approved for the second-line treatment of AIDS-related Kaposi’s sarcoma in 1997.⁴ Recent studies have shown that treatment for various types of advanced solid cancers with paclitaxel and docetaxel often results in the emergence of multi-drug resistance (MDR). This is mainly caused by the over-induction of P-glycoprotein.⁵ Therefore, it is important to explore new taxoids either with improved activity against various classes of tumors, especially against drug-resistant human cancers or as functional inhibitors of the P-glycoprotein. In this letter, we thus report that hydroxylation of the C-1 position of 2-deacetoxytaxinine J derivative **3** has been successfully achieved by a simple chemical oxidation using dimethyldioxirane (DMDO)⁶ leading to the 1 β -hydroxy α -4(20)-epoxide **6** and 1 β -hydroxy β -4(20)-epoxide **7** in our ongoing chemical conversions⁷ of natural taxoids available from Japanese yew for new taxoid anticancer agents for clinical use.

* Corresponding author. Tel/fax: +81-22-717-8783; e-mail: oritani@biochem.tohoku.ac.jp (T. Oritani)



The 2-deacetytaxinine J **2**,⁸ which is a natural taxoid isolated from the Japanese yew *Taxus cuspidata*, shows no cardiac toxicity,⁹ but might thus serve as an important starting material for the synthesis of new functional inhibitors of the P-glycoprotein. Treatment of **2** with hydroxyamine hydrochloride¹⁰ in ethanol under reflux followed by the protection of 5-hydroxy group with chlorotriethylsilane gave the protected taxoid **3**. The epoxidations of taxinine derivatives leading to 4(20)-epoxides using DMDO (1.8 equiv.) within the stereoselectivity of the α : β ratio in a range from 99:1 to 1:4 have been reported.¹¹ However, it was found that treatment of **3** with a large excess of DMDO (15 equiv.) in CH_2Cl_2 at room temperature for 48 h afforded the α -4(20)-epoxide **4** and β -4(20)-epoxide **5** in 60% yield (α : β =1.1:1), and the unexpected two more polar products **6** and **7** in 38% yield which were later determined to be the 1 β -hydroxy α -4(20)-epoxide and 1 β -hydroxy β -4(20)-epoxide,¹² respectively (Scheme 1). While treatment of **3** with 30 equiv. of DMDO under similar conditions gave only **6** and **7** in 90% yield as the reaction products. Additionally, the compounds **4** and **5** could be further converted completely into **6** and **7**, respectively, using 25–30 equiv. of DMDO.



Scheme 1.

The structures and stereochemistries of **4–7** were determined by their NMR data including 2D NMR experiments. The NOESY correlations of H-20a to H₂-2 and H₃-19, plus H-20b to H-5 β and H-6 β were observed in **4** demonstrating the 4(20)-epoxide moiety which possesses the α -orientation. Whereas the β -orientation for the 4(20)-epoxide moiety of **5** was established by the NOESY correlations of H-20a to H-14 α . In comparison to the NMR data of **4**, those of **6** displayed high similar structural signals to **4** except where both H₂-14 showed the doublet–doublet signals at δ_{H} 2.38 ($J=14.6, 9.5$ Hz) and δ_{H} 1.86 ($J=14.6, 5.3$ Hz), respectively, and a new downfield quaternary carbon (C-1) resonanced at δ_{C} 77.0 (s), attributed to the presence of a hydroxy group located at C-1. This was substantiated by IR absorption at 3400 cm^{-1} and HMRS (FAB) giving the molecular formula as $\text{C}_{34}\text{H}_{54}\text{O}_{11}\text{Si}$ for $(\text{M}+\text{Na})^+$, calcd: 689.3329; found: 689.3326. Fortunately, the similar assignment of a hydroxyl group located at C-1 was obtained by comparisons of the NMR data of **7** with those of **5** and **6**. The β -orientation of the 1-hydroxyl group for **6** and **7** was established by NOESY studies in which both compounds showed the absence of NOESY correlations of H-2 β to H-14 β , and H-3 α to H-14 α .¹³

Formation of 1 β -hydroxy 4(20)-epoxides **6** and **7** may be explained via a known 'insertion' of an *O*-atom into an 'unactivated' C–H bond of saturated hydrocarbon as oxygenations of alkanes by metalloporphyrins¹⁴ or by ozone¹⁵ (Fig. 1). This insertion reaction is proposed to involve a transition state I, which might have an appreciable diradical character that leads to the formation of alcohol and it is mediated by caged radical-pairs II; then, in-cage oxidation would lead to the production of alcohols.¹⁶ While the dioxirane O–O bond is being broken, significant widening of the O–C–O angle from 60° to nearly 107° occurs, so that tertiary selectivity asymmetry might serve to relax the energy requirements of the *O*-atom insertion.¹⁷ According to this sequence, the high stereoselectivity recorded might be accommodated, provided one makes the 'assumption' that an in-cage collapse of the radical pair to products occurs faster than the loss of configuration of the caged carbon radical. This stereochemistry of *O*-atom insertion reaction was supported by the regio- and stereospecific hydroxylation of taxoid **3** using DMDO to afford 1 β -hydroxy 4(20)-epoxides **6** and **7** with complete retention of configuration. It is thought that highly regioselective hydroxylation at C-1 is more predominant at C-3 in **3**, attributed to a large steric hindrance at C-3 for an optimum stereoalignment of dioxirane attack owing to the convex shape of the taxane skeleton. In fact, the high regioselectivities of tertiary versus secondary and the complete stereospecificities with retention displayed in the *O*-atom insertion with dioxiranes have been reported.¹⁸

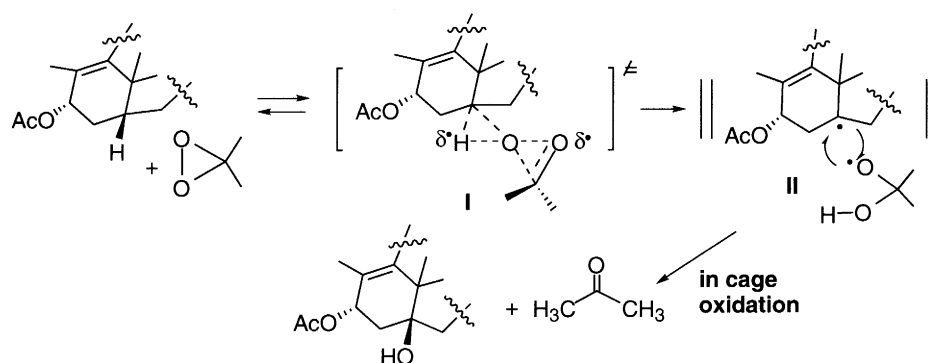


Fig. 1. The proposed mechanism of *O*-atom insertion reaction of taxoid **3** leads to **6** and **7**

Although a selective microbial hydroxylation of taxoid has been recently reported,¹⁹ this microbial enzymatic reaction led to the hydroxylations at the C-1 and C-14 positions to afford the two compounds, 1 β -hydroxy and 14 β -hydroxy derivatives in a 1.5:1 ratio as well as A-ring contracted alcohol. To our knowledge, the above-described high regio- and stereospecific hydroxylation leading to only 1 β -hydroxy with dimethyldioxirane is the first example to date of the introduction of the hydroxy group at C-1 in the taxoid series by a chemical method. It is worthy to note that this chemistry will be applied for the semisynthesis of biologically active 1 β -hydroxy taxoids from a lot of 1 β -hydroxy taxoids contained in most of the *Taxus* species. These applications are in progress with encouraging results by our group.

Acknowledgements

We would like to express our gratitude to Dr. Q.-W. Shi, and Dr. Q. Cheng wishes to thank the Japan Society for the Promotion of Science for a JSPS fellowship.

References

- Wani, M. C.; Taylor, H. L.; Wall, M. E.; Coggon, P.; McPhail, A. T. *J. Am. Chem. Soc.* **1971**, *93*, 2325–2327.
- (a) Bissery, M. C.; Guénard, D.; Guéritte-Vogelein, F.; Lavelle, F. *Cancer Res.* **1991**, *51*, 4845–4848. (b) Guénard, D.; Guéritte-Vogelein, F.; Potier, P. *Acc. Chem. Res.* **1993**, *26*, 160–167.
- Schiff, P. B.; Fant, J.; Horwitz, S. B. *Nature* **1979**, *227*, 665–667.
- Ojima, I.; Wang, T.; Miller, M. L.; Lin, S.; Borella, C. P.; Geng, X.; Pera, P.; Bernacki, R. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 3423–3428.
- Bosch, I.; Croop, J. *Biochem. Biophys. Acta* **1996**, *1288*, 37–54.
- (a) Adam, W.; Bialas, J.; Hadjiarapoglou, L. *Chem. Ber.* **1991**, *124*, 2377. (b) Ferrer, M.; Gilbert, M.; Sanchez-Baeza, F.; Messegue, A. *Tetrahedron Lett.* **1996**, *37*, 3585–3586.
- (a) Cheng, Q.; Oritani, T.; Horiguchi, T. *Tetrahedron*, **1999**, *55*, 12 099–12 108. (b) Cheng, Q.; Oritani, T.; Horiguchi, T.; Yamada, T.; Hassner, A. *Tetrahedron* **2000**, *56*, 1181–1192. (c) Cheng, Q.; Oritani, T.; Horiguchi, T.; Yamada, T.; Mong, Y. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 517–521. (d) Cheng, Q.; Oritani, T.; Horiguchi, T. *Tetrahedron* **2000**, *56*, 1667–1679. (e) Cheng, Q.; Oritani, T.; Horiguchi, T. *Chin. Chem. Lett.* **2000**, *11*, in press.
- Liang, J.-Y.; Min, Z.-D.; Niwa, M. *Acta Chim. Sin.* **1988**, *46*, 1053–1055.
- Alloatti, G.; Penna, C.; Levi, R. C.; Gallo, M. P.; Appendino, G.; Fenoglio, I. *Life Sci.* **1996**, *58*, 845–852.
- Bathini, Y.; Micetich, A. G.; Daneshalab, M. *Synth. Commun.* **1994**, *24*, 1513–1517.
- Hosoyama, H.; Shigemori, H.; In, Y.; Ishida, T.; Kobayashi, J. *Tetrahedron* **1998**, *54*, 2521–2528.
- Selected spectroscopic data of **6** and **7** are as follows. 1 β -OH- α -4(20)-epoxide **6**: [α]_D²³ +27.4 (*c* 0.12, CHCl₃), ¹H NMR (500 MHz, CDCl₃) δ : 6.24 (d, 1H, *J*=11.2 Hz), 5.90 (d, 1H, *J*=11.2 Hz), 5.91 (m, 1H), 5.64 (dd, 1H, *J*=11.7, 4.8 Hz), 3.35 (brt, 1H), 2.77 (d, 1H, *J*=6.4 Hz), 2.56 (d, 1H, *J*=4.2 Hz), 2.55 (d, 1H, *J*=4.2 Hz), 2.38 (dd, 1H, *J*=14.6, 9.5 Hz), 2.15 (s, 3H), 2.14 (s, 3H), 2.07 (s, 3H), 2.04 (s, 3H), 1.99 (m, 1H), 1.97 (s, 3H), 1.86 (dd, 1H, *J*=14.6, 5.3 Hz), 1.84 (m, 1H), 1.63 (m, 1H), 1.59 (s, 3H), 1.23 (s, 3H), 1.10–0.94 (m, 10H), 0.87 (s, 3H), 0.73–0.63 (m, 6H). ¹³C NMR (125 MHz, CDCl₃) δ : 171.5 (s), 171.1 (s), 170.4 (s), 169.8 (s), 141.1 (s), 134.0 (s), 77.0 (s), 76.9 (d), 73.0 (d), 71.9 (d), 71.4 (s), 70.1 (s), 60.0 (s), 48.8 (t), 47.6 (s), 44.8 (s), 41.1 (t), 37.5 (t), 36.3 (d), 32.6 (t), 28.0 (q), 23.1 (q), 22.2, 22.1, 21.7, 21.6 (4 \times q), 15.4 (q), 14.0 (q), 6.9 (q), 4.7 (t). 1 β -OH- β -4(20)-epoxide **7**: [α]_D²³ +53.7 (*c* 2.0, CHCl₃), ¹H NMR (500 MHz, CDCl₃) δ : 6.19 (d, 1H, *J*=11.2 Hz), 6.07 (m, 1H), 5.87 (d, 1H, *J*=11.2 Hz), 5.59 (dd, 1H, *J*=10.7, 4.6 Hz), 3.34 (t, 1H, *J*=3.4 Hz), 2.85 (d, 1H, *J*=3.6 Hz), 2.69 (dd, 1H, *J*=4.9, 2.2 Hz), 2.48 (dd, 1H, *J*=14.9, 9.5 Hz), 2.43 (d, 1H, *J*=3.6 Hz), 2.12 (s, 6H), 2.07 (s, 3H), 2.04 (s, 3H), 2.02 (m, 1H), 1.98 (s, 3H), 1.73–1.70 (m, 2H), 1.63 (dd, 1H, *J*=14.9, 5.1 Hz), 1.57 (s, 3H), 1.19 (s, 3H), 1.09 (s, 3H), 1.00–0.94 (m, 9H), 0.83 (dd, 1H, *J*=15.4, 2.2 Hz), 0.68–0.60 (m, 6H). ¹³C NMR (125 MHz, CDCl₃) δ : 171.3 (s), 171.1 (s), 170.4 (s), 169.9 (s), 140.7 (s), 135.1 (s), 76.9 (d), 76.5 (s), 73.8 (d), 72.0 (d), 71.5 (s), 70.1 (s), 61.5 (s), 48.6 (t), 47.2 (s), 44.9 (s), 40.6 (t), 37.2 (t), 36.1 (d), 32.4 (t), 28.3 (q), 22.5 (q), 22.1, 22.0, 21.7, 21.6 (4 \times q), 18.1 (q), 15.7 (s), 6.7 (q), 4.6 (t).
- The comparisons of the NOE data of **6** and **7** with those of the closely related natural and semisynthetic 1 β -OH-taxoids exhibit if the 1-hydroxy is α -orientated; there should be strong NOE relationships between H-2 β and H-14 β , and between H-3 α and H-14 α . These are confirmed by the molecular model studies. In fact, to our knowledge, so far all natural and semisynthetic 1-OH taxoids having been reported are β -orientated. In addition, it is unreasonable to think about a very steric 1- α -hydroxyl form in a rigid taxane skeleton by an analysis of molecular model.
- Hill, C. L. In *Advances in Oxygenated Processes*, Baumstark, A. L., Ed.; JAI: Greenwich, CT, 1988; pp. 1–30.
- Giamalva, D. H.; Church, D. F.; Pryor, W. A. *J. Org. Chem.* **1988**, *53*, 3429–3432.
- Once a radical-pair is produced from state I, both radicals might diffuse out of the cage. However, the formation of a pair of separate radicals can be rejected because of the observed complete retention of configuration and no products derived from free radical reactions, or even arising from the radical-initiated chain decomposition of dioxirane itself; these corroborate that discrete free radicals are not involved in this *O*-atom insertion.
- Bach, R. D.; Andres, J. L.; Owensby, A. L.; Schlegel, H. B.; McDouall, J. J. W. *J. Am. Chem. Soc.* **1992**, *114*, 7207–7217.
- (a) Murray, R. W.; Jeyaraman, R.; Mohan, L. *J. Am. Chem. Soc.* **1986**, *108*, 2470–2472. (b) Mello, R.; Cassidei, L.; Fiorentino, M.; Fusco, C.; Curci, R. *Tetrahedron Lett.* **1990**, *31*, 3067–3070. (c) Adam, A.; Asensio, G.; Curci, R.; Gonzales-Nunez, M. E.; Mello, R. *J. Org. Chem.* **1992**, *57*, 953–955. (d) Curci, R.; Dinoi, A.; Rubino, M. F. *Pure Appl. Chem.* **1995**, *67*, 811–822.
- Hu, S.; Sun, D.; Tian, X.; Fang, Q. *Tetrahedron Lett.* **1997**, *38*, 2721–2724.